

INHABITANTS OF ENDOPHYTIC *PHOMA*SPP.**Garima Madhariya**

Department of Biotechnology,
Govt. Digvijay Autonomous
P.G. College, Rajnandgaon,
Chhattisgarh, India

Shweta Singh Chauhan

Department of Biotechnology,
Govt. Digvijay Autonomous
P.G. College, Rajnandgaon,
Chhattisgarh, India

Pramod Kumar

Department of Biotechnology.
Govt. Digvijay Autonomous
P.G. College, Rajnandgaon,
Chhattisgarh, India

Shriram Kunjam

Department of Botany,
Govt. V.Y.T. Autonomous P.G.
College, Durg, Chhattisgarh,
India

ABSTRACT

The fungal genus *Phoma* has been documented as exhibiting phytopathogenic properties, as well as functioning as saprophytes in soil. Additionally, this genus has been observed in various environments, including aquatic and aerial settings, marine environments, and as entomopathogens. The taxonomic classification *Phoma* pertains to pycnidia that bear single-celled, transparent conidia and are found on herbaceous stems. *Phoma* spp. had been classified with the class Coelomycetes because they have certain defining physical characteristics. Economically significant crop plants are frequently infected by fungi of the genus *Phoma*. Some *Phoma* species are host specific like *Phoma caloplacae* in *Triticumaestivum* and *Phoma multiristrata* have been found in weed plants *T. procumbens*. Some *Phoma* spp. found in more than plants like *Phomaherbarum*, *Phoma glumarata*, *Phoma enpyrenahabitats* in evergreen tree, mangrove tree, perennial trees, Herbaceous and shrubs also. *Phoma* spp. have the potential to act as opportunistic pathogens for humans, animals, and plants. Many unique and natural products with diverse biological activity have been traced back to *Phoma*, which has gained many prominences. *Phoma* spp. has been found to produce a variety of novel secondary metabolites that exhibit antimicrobial, anti-inflammatory, bio-herbicidal, antiangiogenic, cytotoxic, and anti-HIVactivity.

Keywords: Coelomycetes; Inhabitant; *Phoma* spp.; Pycnidia of *Phoma* spp.; Secondary metabolites.

INTRODUCTION

Phoma is a phylum Ascomycota member and a highly polyphyletic genus with ambiguous species boundaries. Due to these physical characteristics, *Phoma spp.* have traditionally been placed in the class Coelomycetes. Using morphological, more than 220 species were formally recognised in the handbook "Phoma Identification Manual" by Boerema et al. (Boerema et al., 2004). *Phoma spp.* was conceptualized in the 19th century by Italian mycologist Pier Andrea Saccardo 1880 (Kwong-Chung et al., 1992).

The genus *Phoma* fungi are known to undergo asexual reproduction, and their colonies are characterized by a velvety texture that may exhibit a slight powdery appearance, contingent upon the specific species. Pycnidia are characterized by the presence of pycn(idi)ospores or conidia, which are masses of spores consisting of a single cell. The culture characteristics and morphology are subject to variation due to various factors such as pH, temperature, and light, among others (Rajak et al, 1983). *Phoma* species can be distinguished from other dematiaceous fungus by their distinctive morphological characteristics, such as chlamydospores, conidia, and pycnidial conidiomata (fruiting structures that serve as a mechanism of disseminating conidia) (Rai et al., 2014).

Phoma spp. has been observed in various natural habitats, such as aquatic environments, water distribution systems, soil, and air, owing to the widespread prevalence of fungi (Bennett et al., 2018). Water system infiltration by *Phoma spp.* is widely known, and its presence is a public health concern (Babić et al., 2017). *Phoma* species have the ability to transition from being opportunistic microorganisms to pathogenic agents upon encountering a suitable host.

Inhabitants-

Phoma spp. has been reported in multiple natural habitats due to the ubiquitous nature of fungi. The global distribution of *Phoma spp.* in food sources exhibits variability in its levels of exposure. *Phoma spp.* has wide range of host plant and it inhabitate more than one type of host plant and some are document as host specific viz. *P. macrostoma* was observed to specifically inhibit dicot plants' growth (Bailey et al., 2013). *Phoma betae* is endophyte of Desert Plant, Flowering plant ,Herbaceous, perennial as well as in Endangered tree (Parismita et al. 2016). *Phomacaloplaceae*, D. Hawksw. Found in *Triticum aestivum*. Some other *Phoma spp.* have been also found in *Poacea* family crop like *Phoma glomerata*, (A.C.J. Corda) Wollenw. & Hochapfel (Crous et al. 1995).

Table 1 -Biodiversity of Endophytic *Phoma*

S. N o.	Endophytic <i>Phoma</i>	Host flora	Family of the host flora	Type of the host flora	Reference
1	<i>Phoma Americana</i>	<i>Gramineae</i>	Poaceae	Grass	Boerema, 1993
		<i>Gossypium hirsutum</i>	Malvaceae	Cotton	Ek–Ramos et al., 2013
2	<i>Phoma bellidis</i>	<i>Eleocharis dulcis</i>	Cyperaceae	perennial herbaceous plant	Lv et al., 2011
		<i>Ttricyrtis maculate</i>	Liliaceae	Perennial herbaceous	Wang et al., 2019
3	<i>Phoma betae</i>	<i>Beta vulgaris</i>	Amaranthaceae	herbaceous biennial or, rarely, perennial plan	Garibaldi et al., 2007
		<i>Ginkgo biloba</i>	Ginkgoopsida	Endangered tree	Kumaran et al., 2012
		<i>Kalidium foliatum</i> (Pall.) Desert Plant from West China	Amaranthaceae	Desert Plant	Tan et al., 2018
		<i>Artemisia nilagirica</i>	Asteraceae	Herbaceous perennial, aromatic	Parismita et al., 2016
		<i>Arabidopsis thaliana</i>	Brassicaceae	Flowering Weed/ model plant for plant biology and genetics	Junker et al., 2012
4	<i>Phoma cava</i>	<i>Acer saccharum</i>	Sapindaceae	Sugar Maple flowering plant	Vujanovic & Brisson 2002
		<i>Hancornia speciosa</i>	Apocynaceae	Fruit tree	Chagas et al., 2017

		<i>Quercus cerris</i>	Fagaceae	Oak	Ragazzi et al., 1999; Moricca et al., 2012
5	<i>Phoma chenopodiicola</i>	<i>Celtis occidentalis</i>	Cannabaceae	Deciduous tree	Novas & Carmarán 2008
6	<i>Phoma chrysanthemicola, Hollós</i>	<i>Kigelia pinnata</i>	Bignoniaceae	Flowering tree	Maheswari & Rajagopal 2013
		<i>Calotropis procera</i>	Apocynaceae	Shrub	Khan et al., 2007
		<i>Artemisia nilagirica</i>	Asteraceae	Herbaceous perennial, aromatic	Parismita et al., 2016
		<i>Avicennia marina</i> Leaf	Acanthaceae	Mangrove	Bharathidasan & Panneerselvam 2011
7	<i>Phoma cucurbitacearum</i>	<i>Taraxacum coreanum</i>	Asteraceae	Perennial herbaceous	Paul et al., 2006
		<i>Glycyrrhiza glabra</i> Linn.	Fabaceae	Perennial herbaceous legume	Arora et al., 2016
8	<i>Phoma destructive</i>	<i>Gossypium spp.</i>	Malvaceae	Shrub flowering plant	Vieira et al., 2011
9	<i>Phoma draconis</i>	<i>Panax notoginseng</i>	Araliaceae	Medicinal Herb	Zheng et al., 2017
10	<i>Phoma epicoccinia</i>	<i>Phyllanthus amarus</i>	Phyllanthaceae	Tropical coastal herb	Kandasamy et al., 2015
11	<i>Phoma eupatorii</i>	<i>Eupatorium caunabinum</i>	Asteraceae	Herbaceous	De Vries et al., 2018

12	<i>Phoma eupyrena</i>	<i>Coffea arabica</i>	Rubiaceae	Shrub	Oliveira et al., 2014;
		<i>Laguncularia racemosa</i>	Combretacea e	Mangrove	Costa et al., 2012
		<i>Gossypium spp.</i>	Malvaceae	Shrub flowering plant	Vieira et al., 2011
		<i>Clerodendron sp.</i>	Lamiaceae sub family Ajugoideae	Tropical, warm temperate Shrub	Suryan et al., 2016
		<i>Artemisia nilagirica</i>	Asteraceae	Herbaceous perennial, aromatic	Myrchiang et al., 2014; Parismita et al., 2016
13	<i>Phoma exigua, Desmazières, J.B.H.J.</i>	<i>Calotropis gigantea L.</i>	Apocynaceae	Shrub	Selvanatha n et al., 2011
		<i>Coffea arabica L.</i>	Rubiaceae	Shrub	Fernandes et al., 2009
		<i>Ensete ventricosum</i>	Musaceae	Evergreen Herbaceous	Chauhan et al., 2019
		<i>Artemisia nilagirica</i>	Asteraceae	Herbaceous perennial, aromatic	Parismita et al., 2016
		<i>Ptychosperma macarthurii</i>	Arecaceae	Palm	Song et al., 2015
		<i>Quercus ilex L.</i>	Fagaceae	Evergreen oak	Fisher et al., 1994
14	<i>Phoma fimetii, Brunaud</i>	<i>Vulpia ciliata spp. ambigua</i>	Poaceae subfamily Pooideae	Grass	Newsham, 1994
		<i>Eucalyptus globules</i>	Myrtaceae	Evergreen tree	Simeto et al., 2005

15	<i>Phoma gardenia</i>	<i>Brugmansia aurea</i> Lagerh	Solanaceae	Sub-tropical ornamental shrub,	Singh et al., 2017
16	<i>Phoma glomerata</i> , (A.C.J. Corda) Wollenw. & Hochapfel	<i>Acer truncatum</i>	Sapindaceae	Deciduous tree	Sun et al., 2011
		<i>Panicum virgatum</i> L.	Poaceae	Perennial grass	Ghimire et al., 2011
		<i>Vitis vinifera</i> Grapes	Vitaceae	Liana	González & Tello 2011
		<i>Sporobolus pyramidatus</i> (Lam.) Hitchc	Poaceae	Annual grass	Loro et al., 2012
		<i>Pappophorum krapovickasii</i> Roseng.	Poaceae sub family Chloridoidea e	Perennial grass	
		<i>Cyperus laevigatus</i> L.	Cyperaceae	Perennial sedge	
		<i>Trachycarpus fortunei</i>	Arecaceae	Palm	Taylor et al., 1999
		<i>Cucumis sativus</i>	Cucurbitaceae	Vegetable	Waqas et al., 2012
		<i>Glycine max</i> From leaves	Fabaceae Sub family Faboideae	Edible bean crop	Fernandes et al., 2015
		<i>Triticum aestivum</i>	Poaceae	Cereal crop	Crous et al., 1995
		<i>Gossypium spp.</i>	Malvaceae	Shrub flowering plant	Vieira et al., 2011
		<i>Salvia miltiorrhiza</i>	Lamiaceae	Deciduous perennial herb	Li et al., 2016

17	<i>Phoma hedericola</i> , (Durieu & J.P.F.C. Montagne) Boerema	<i>Avicennia marina</i> Leaf	Acanthaceae	Mangrove	Bharathida san & Panneersel vam 2011
		<i>Menthe viridis</i> Stem	Lamiaceae	Perennial herb, medicinal, food supp.	Kumar et al., 2016
		<i>Ricinus communis</i> <i>Linn.</i> Leaf	Euphorbiace ae Sub family Acalyphoide ae	Perenial flowering, suckering shrub	Sandhu et al., 2014
		<i>Calotropis procera</i>	Apocynaceae	Shrub	Juyal et al., 2017 and Khan et al., 2007
18	<i>Phoma herbarum</i> , G.D. Westendorp	<i>Rauwolfia serpentine</i>	Apocynaceae	Perennial shurb	Das & Chatterjee (2023).
		<i>Holcus lanatus</i>	Poaceae	Perennial grass	Márquez et al., 2010
		<i>Pinus halepensis</i>	Pinaceae	Cone Pinus tree	Botella & Diez 2011
		<i>Olea europaea L.</i>	Oleaceae	Evergreen small, tree	Materatski et al., 2019
		<i>Laguncularia racemosa</i>	Combretacea e	Mangrove	Costa et al., 2012
		<i>Curcuma longa L.</i>	Zingiberacea e	Perennial rhizomatous herbaceous	Gupta et al., 2016
		<i>Taxus x media</i>	Taxaceae	Conifer	Xiong et al., 2013
		<i>Coffea arabica L.</i>	Rubiaceae	Shrub	Fernandes

				et al., 2009	
	<i>Helianthemum guerrae</i>	Cistaceae	Shrub	Macia-Vicente et al., 2008	
	<i>Piper hispidum</i> Sw	Piperaceae	Perennial shrub	Orlandelli et al., 2012	
	<i>Salvia miltiorrhiza</i>	Lamiaceae	Deciduous perennial herb	Chen et al., 2020	
19	<i>Phoma jolyana</i> , Piroz. & Morgan-Jones	<i>Eucalyptus nitens</i>	Myrtaceae	Tall tree	Smith et al., 1996
		<i>Forsythia giraldiana</i>	Oleaceae	Deciduous shrub	Sun et al., 2008
		<i>Azadirachta indica</i>	<u>Meliaceae</u>	evergreen tree	Iqbal et al., 1984
20	<i>Phoma leonuri</i> , Letendre	<i>Phoradendron perrottettii</i>	Santalaceae		De Abreu et al., 2010
21	<i>Phoma leveillei</i> , Boerema & G.J. Bollen	<i>Arabidopsis thaliana</i>	Brassicaceae	Flowering Weed/ model plant for plant biology and genetics	Junker et al., 2012
		<i>Rhizophora mucronata</i>	Rhizophoraceae	Tropical Mangrove	Humzah et al., 2018
		<i>Quercus ilex L.</i>	Fagaceae	Evergreen oak	Fisher et al., 1994
		<i>Cantharanthus roseus</i>	Apocynaceae	Evergreen Sub-shrub or herb	Krishnamurthy et al., 2008
		<i>Rauwolfia serpentine</i>	Apocynaceae	Perennial shurb	
		<i>Helianthemum guerrae</i>	Cistaceae	Shrub	Macia-Vicente et al., 2008
		<i>Ononis natrix</i>	Fabaceae	Perennial shrubby	

		<i>subsp. Ramosissima</i>			
		<i>Euphorbia sancte Catherine</i>	Euphorbiaceae	Aromatic herb	Selim et al., 2018
22	Phoma <i>lingam</i> , (Tode) Desmazières, J.B.H.J.	<i>Eucalyptus globulus</i>	Myrtaceae	Evergreen, tall tree	Bettucci & Saravay 1993
		<i>Pinus tabulaeformis</i> Mountain	Pinaceae	Pinus	Guo et al., 2008
		<i>Prosopis juliflora</i> (S.W.) DC. Leaves	Fabaceae	Weed, Shrub or small tree	Srivastava and Anandrao, 2015
23	<i>Phoma macrostoma</i> , J.P.F.C. Montagne				
		<i>Arabidopsis thaliana</i>	Brassicaceae	Flowering Weed/ model plant for plant biology and genetics	García et al., 2013
		<i>Olea europaea L.</i> Leaves, oil tree Mediterranean	Oleaceae	Evergreen small, tree	Materatski et al., 2019
24	<i>Phoma medicaginis</i> , Malbranche & Roumeguère	<i>Glycyrhiza glabra</i> Linn.	Fabaceae	Herb, perennial, legume	Nalli et al., 2019
		<i>Medicago sativa</i>	Fabaceae	Perennial flowering legume	Weber et al., 2004
		<i>Medicago lupulina</i>	Fabaceae Sub family Faboideae	Annual or shot-live perennial legume	
		<i>Aquilaria sinensis</i> China	Thymelaeaceae	Evergreen tree	Cui et al., 2011
		<i>Panicum virgatum</i> L.	Poaceae	Perennial grass	Ghimire et al., 2011
		<i>Taxus globosa</i>	Taxaceae	Evergreen Shrub	Rivera–Orduña et

					al., 2011
	<i>Vitis vinifera</i> Grapes	Vitaceae	Liana		Cosoveanu et al., 2014
	<i>Mikania cordata</i>	Asteraceae	Creeping, perennial		Jayatiake and Munasinghe, 2020
	<i>Boswellia</i>	Burseraceae	Moderate size tree and shrub		Khan et al., 2016
	<i>Taxus wallichiana</i> var. <i>mairei</i>	Taxaceae	Deciduous/ evergreen shrub		Zaiyou et al., 2017
25	<i>Phoma microchlamydospora</i> , Aveskamp & G.J.M. Verkley	<i>Terminalia mantaly</i>	Combretaceae	Evergreen small to medium tree	Toghueo et al., 2017
26	<i>Phoma moricola</i> , P.A. Saccardo	<i>Paspalum ligulare</i> Nees	Poaceae	Herb	Loro et al., 2012
	<i>Eremophila longifolia</i> Australia	Scrophulariaceae	Shrub or small flowering tree		Zaferanloo et al., 2018
	<i>Solanum cernuum</i> Vell.	Solanaceae	Shrub or small tree		Vieira et al., 2012
	<i>Garcinia parvifolia</i>	Clusiaceae	Tropical evergreen tree		Sim et al., 2010
	<i>Echinacea purpurea</i> (L.) Moench	Asteraceae	Perennial, herbaceous		Luiz et al., 2012
27	<i>Phoma multirostrata</i> , (P.N. Mathur,	<i>Trachycarpus fortunei</i> (Temperate palm)	Arecaceae	Palm	Taylor et al., 1999

	S.K. Menon & Thirum.) Dorenb. & Boerema	<i>Cantharanthus roseus</i> <i>Withinia somnifera</i> <i>Cassia tora</i> <i>Securinega suffruticosa</i> <i>Eupatorium adenophorum</i> Leaf <i>Centella asiatica</i> Leaves, root, stolons <i>Cantharanthus roseus</i> Leaves	Apocynaceae Solanaceae Fabaceae Phyllanthaceae Asteraceae Apiaceae Apocynaceae	Evergreen Sub-shrub or herb Perennial shrub Annual Herb Deciduous shrub Perennial, herb Perennial, flowering herb Evergreen Sub-shrub or herb	Krishnamurthy et al., 2008 Du et al., 2020 Chen et al., 2015 Radiastuti et al., 2019 Khiralla et al., 2016
28	<i>Phoma nebulosa</i> , (Persoon) Berkeley	<i>Cirsium arvense L.</i> <i>Mimusops elengi</i> Leaf <i>Trachycarpus fortunei</i>	Asteraceae Sapotaceae Arecaceae	Herb, perennial C3 carbon fixing plant Tropical evergreen tree Palm	Schulz et al., 1998 Kannan et al., 2017 Taylor et al., 1999
29	<i>Phoma pinodella</i> , (L.K. Jones) Morgan-Jones & K.B. Burch	<i>Hevea brasiliensis</i> Rubber plant <i>Trachycarpus fortunei</i>	Euphorbiaceae Arecaceae	Deciduous tree Palm	Gazis & Chaverri 2010 Taylor et al., 1999
30	<i>Phoma pomorum</i> , Thümen, F. von.	<i>Glycine max</i> <i>Salsola collina</i> <i>Suaeda salsa</i>	Fabaceae Sub family Faboideae Chenopodiaceae Amaranthace	Edible bean crop Desert weed plant Annual herb	Fernandes et al., 2015 Sun et al., 2012

			ae		
		<i>Trachycarpus fortunei</i>	Arecaceae	Palm	Taylor et al., 1999
		<i>Citrus limon</i>	Rutaceae	Small evergreen plant	Douanla-Meli et al., 2013
31	<i>Phoma putaminum</i> , C. Spegazzini	<i>Dendrobium crumenatum</i>	Orchidaceae Subfamily - Epidendroideae	Pigeon orchid, large but short-lived, strongly scented white flowers	Maciá-Vicente et al., 2008
		<i>Bauhinia forticata</i>	Fabaceae	Brazilian orchid tree, flowering tree in the pea family	Bezerra et al., 2015
		<i>Dysosma versipellis</i>	Berberidaceae	flowering barberry plant	Tan et al., 2018
32	<i>Phoma radicina</i> , (McAlpine) Boerema	<i>Panax ginseng Meyer</i>	Araliaceae	Medicinal Herb	Park et al., 2012
		<i>Panax notoginseng</i>	Araliaceae	Medicinal Herb	Zheng et al., 2017
		<i>Salvia abrotanoides</i>	Lamiaceae	Russian sage, flowering herb	Teimoori-Boghsani et al., 2020
		<i>Armoracia rusticana</i>	Brassicaceae	Perennial flowering plant	Szűcs et al., 2018
33	<i>Phoma selaginellicola</i> , Gruyter, Noordeloos, Aa & Boerema	<i>Dysosma versipellis</i> Root	Berberidaceae	Flowering barberry plant	Tan et al., 2018b
		<i>Acer negundo L.</i> Root	<u>Sapindaceae</u>	Fast-growing, short-lived tree with opposite, compound leaves	Bukharina et al., 2018

		<i>Thymus eriocalyx</i>	Lamiaceae	Flowering herb	Masumi et al., 2014
34	<i>Phoma solani</i> , Cooke & Harkn.	<i>Boswellia sacra</i>	<u>Burseraceae</u>	Small <u>deciduous</u> tree	El– Nagerabi et al., 2014
35	Phoma sorghina, (P.A. Saccardo) Boerema, Dorenb. & Kesteren	<i>Chloris inflate</i> (=C. barbata Sw.)	Poaceae	Monocot weed	Loro et al., 2012
		<i>Ziziphus spinachristi</i>	Rhamnaceae	Evergreen tree, arid and semi-arid	El– Nagerabi et al., 2013
		<i>Brachiaria</i>	<u>Poaceae</u>	Grass	Gama et al., 2019
		<i>Musa spp</i> Leaves	<u>Musaceae</u>	Largest herbaceous plants	Zakaria and Aziz 2018
36	Phoma terrestris, H.N. Hansen (Present Name : Setophoma terrestris)	<i>Mimosops elengi</i>	<u>Sapotaceae</u>	Evergreen tree, tropical forests	Deshmukh et al., 2009
		<i>Panax ginseng</i> Meyer	<u>Araliaceae</u>	Perennial, medicinal plant	Park et al., 2015
		<i>Brassica oleracea</i> var. <i>acephala</i>	<u>Brassicaceae</u>	Food <u>crop</u> plant	Poveda et al., 2020
37	Phoma tropica, R. Schneid. & Boerema	<i>Opuntia ficus-indica</i> Mill.	Cactaceae	<u>Cactus</u> <u>arid</u> and <u>semiarid</u>	Bezzera et al., 2012
		<i>Fucus spiralis</i> <u>seaweed</u> , a <u>brown</u> Algae	<u>Fucaceae</u>	<u>Seaweed</u> , a <u>brown</u> Algae	Osterhage et al., 2002
38	Phoma sp.	<i>Calotropis procera</i>	Apocynaceae	Shrub	Khan et al.,

					2007
39	Phoma sp.	<i>Tinospora cardifolia</i>	Menisperma ceae	Herbaceous vine	Kedar et al., 2014
40	Phoma sp.	<i>Calotropis procera</i>	Apocynaceae	Flowering plant	Kedar et al., 2014
41	Phoma sp.	<i>Taxus wallichinia</i> ,	<u>Taxaceae</u>	<u>Evergreen coniferous tree</u>	Yang et al., 1994
42	Phoma sp.	<i>Larrea tridentata</i> Desert	<u>Zygophyllaceae</u>	<u>Flowering plant</u>	Strobel et al., 2011
43	Phoma sp. NRRL 46751	<i>Saurauia scaberrinae</i>	Actinidiaceae	<u>Flowering plant</u>	Wijeratne et al., (2013)
44	Phoma sp. EL002650	<i>Artemisia princeps</i> <i>Pamp</i>	<u>Asteraceae</u>	<u>Perennial</u>	Kim et al., 2018
45	Phoma sp. YUD17001	<i>Gastrodia elata</i>	Orchidaceae	Perennial herb	Li et al., 2020
46	Phoma sp. ZJWCF006	<i>Arisaema erubescenes</i>	<u>Araceae</u>	Flowering plant	Wang et al., 2012
47	Phoma sp.	<i>Artemisia thuscula</i>	<u>Asteraceae</u>	Evergreen shrub	Cosoveanu et al., 2018
48	Phoma sp.	<i>Sapindus saponaria</i> L.	<u>Sapindaceae</u> Ranunculaceae	Small to medium-sized <u>deciduous tree</u>	Garcia et al., 2012
49	Phoma sp.	<i>Aconitum vilmorinianum</i>	Ranunculaceae	Herbaceous perennials or biennials	Liu et al., 2019
50	Phoma sp.	<i>Artemisia annua</i>	<u>Asteraceae</u>	Annual short-day plant	Wati et al., 2019
51	Phoma sp.	<i>Rhizophora mucronata</i>	Rhizophoraceae	Small to medium size evergreen tree	Hamzah et al., 2018
52	Phoma sp.	<i>Facus serratus</i>	Fucaceae	Robust alga	Hussain et al., 2014

53	Phoma sp.	<i>Luehea divaricata</i>	<u>Malvaceae</u>	<u>Medicinal plant</u>	Alberto et al., 2016
		<i>Sapindus saponaria</i>	Sapindaceae	Small to medium-sized <u>deciduous tree</u>	
54	Phoma sp. LK13	<i>Moringa peregrine</i>	<u>Moringaceae</u>	Flowering plant	Khan et al., 2014
55	Phoma sp.	<i>Rhododendrons</i>	Ericaceae	Either <u>evergreen</u> or <u>deciduous shrubs</u>	Purmale et al., 2012
56	Phoma sp. PT01	<i>Mitrajyna javanica Koord and val</i>	Rubiaceae		Pharamat et al., 2013
57	Phoma sp.	<i>Calotropis gigantean</i>	Apocynaceae	Waxy flowers <u>shrubs</u>	Hemamalini et al., 2015
58	Phoma sp.	<i>Ficus infectoria</i>	<u>Moraceae</u>	<u>Semi-evergreen tree</u>	Rajagopal et al., 2008
		<i>Ficus microcarpa</i> (Tropical forest western ghats)	Moraceae	Tropical ornamental tree	
59	Phoma sp.	<i>Musa acuminata</i> , (North queensland)	<u>Musaceae</u>	<u>Evergreen perennial</u> , not a <u>tree</u>	Brown et al., 1998
60	Phoma sp.	<i>Rhapis humulis</i>	Arecaceae	Evergreen shrub	Jiao jiao et al., 2016
		<i>Wodyetia bifurcate</i>	Arecaceae	Small or medium-sized, solitary palm	
		<i>Chrysalidocarpus lotescens</i>	<u>Arecaceae</u>	<u>Flowering plant</u>	
		<i>Veitchia merrillii</i>	<u>Arecaceae</u>	Tropical palm tree	
		<i>Phoenix roebelenii</i>	Arecaceae	Slow-growing slender tree	
61	Phoma sp. EA-122	<i>Eupatorium adenop horum</i>	Asteraceae	Subtropical and warmer temperate	Chen et al., 2015
62	Phoma sp.	<i>Hyparrhenia hirta</i>	<u>Poaceae</u>	Tenacious <u>noxious weed</u>	White and Backhouse., 2007
		<i>Botriochloa macra</i>	<u>Poaceae</u>	Perennial grass	

					Backhouse, , 2007
63	Phoma sp. SYSU-SK-7	<i>Kandelia candel</i>	Rhizophorac eae	Shrub or small tree up to 10 metres (Mangroove)	Chen et al., 2019
64	Phoma sp.	<i>Triticum aestivum</i> <i>Leaf</i>	Poaceae	Edible Crop	Larran et al., 2002

Endophytes play a significant role in drug discovery because the interaction between host plants and endophytes enhances the possibility of finding new natural products (Szűcs et al., 2018). According to this review phoma species produces some metabolites such as Polyketide, Taxol/Spiciferon 1-7, Thiodiketopiperazine , Salvianolic Acid C, Gentisyl alcohol/Tanshinone, Macrophin, Brefeldin A/Glucosidases/Paclitaxel, amylase/cellulose/lipase, Cytochalasins, Cryptotanshinone/ Glucosinolate/isothiocyanate, Amylase, Ergoflavin, 5-hydroxyramulosin7-methoxycoumarin, Alkaloids, Isochromanone derivatives, Phenolic compounds/aliphatic easter derivatives/ fatty acids, Polyketides, 14-nordrimane produced by *Phoma bellidis*, *Phoma betae*, *Phoma cucurbitacearum*, *Phoma glomerata*, *Phoma herbarum*, *Phoma macrostoma*, *Phoma medicaginis*, *Phoma microchlamydospora*, *Phoma multirostrata*, *Phoma radicina*, *Phoma selaginellicola*, *Phoma terrestris*, *Phoma tropica*, *Phoma* sp. NRRL 46751, *Phoma* sp. EL002650, *Phoma* sp. YUD17001, *Phoma* sp. SYSU-SK-7, *Phoma* sp respectively. These metabolites have antimicrobial, antiviral, antifungal, antibacterial, cytotoxic, anti-cancer, anti-proliferative, antitubercular activity, which have been proved by various studies.

CONCLUSION

The taxonomic classification of *Phoma* spp. is still a subject of controversy within academic circles, with both morphological and molecular data being utilised in attempts to resolve the issue. The taxonomic classification of the *Phoma* genus remains a subject of controversy despite the utilisation of both molecular and morphological data. There have been reports of perplexing and intersecting characters observed among various *Phoma* species belonging to identical sections. Several studies have been conducted regarding the cultural and morphological characterization of *Phoma* species in India. However, no comprehensive research has been conducted on the development of identification keys for a particular taxonomic group.

REFERENCES

1. Alberto, R. N., Costa, A. T., Polonio, J. C., Santos, M. S., Rhoden, S. A., Azevedo, J. L. D., & Pamphile, J. A. (2016). Extracellular enzymatic profiles and taxonomic identification of endophytic fungi isolated from four plant species. *Genet Mol Res*, 15(4), gmr15049016.
2. Arora, P., Wani, Z. A., Nalli, Y., Ali, A., & Riyaz-Ul-Hassan, S. (2016). Antimicrobial Potential of Thiodiketopiperazine Derivatives Produced by Phoma sp., an Endophyte of *Glycyrrhiza glabra* Linn. *Microbial Ecology*, 72, 802-812.
3. Bailey, K. L., Falk, S., Derby, J. A., Melzer, M., & Boland, G. J. (2013). The effect of fertilizers on the efficacy of the bioherbicide, *Phoma macrostoma*, to control dandelions in turfgrass. *Biological Control*, 65(1), 147-151.
4. Bettucci, L., & Saravay, M. (1993). Endophytic fungi of *Eucalyptus globulus*: a preliminary study. *Mycological Research*, 97(6), 679-682.
5. Bennett, A., Ponder, M. M., & Garcia-Diaz, J. (2018). Phoma infections: classification, potential food sources, and their clinical impact. *Microorganisms*, 6(3), 58.
6. Bezerra, J. D. P., Santos, M. G. S., Svedese, V. M., Lima, D. M. M., Fernandes, M. J. S., Paiva, L. M., & Souza-Motta, C. M. (2012). Richness of endophytic fungi isolated from *Opuntia ficus-indica* Mill.(Cactaceae) and preliminary screening for enzyme production. *World Journal of Microbiology and Biotechnology*, 28, 1989-1995.
7. Bezerra, J. D., Nascimento, C. C., Barbosa, R. D. N., da Silva, D. C., Svedese, V. M., Silva-Nogueira, E. B., ... & Souza-Motta, C. M. (2015). Endophytic fungi from medicinal plant *Bauhinia forficata*: Diversity and biotechnological potential. *Brazilian Journal of Microbiology*, 46, 49-57.
8. Bharathidasan, R., & Panneerselvam, A. (2011). Isolation and identification of endophytic fungi from *Avicennia marina* in Ramanathapuram District, Karankadu, Tamilnadu, India. *European Journal of Experimental Biology*, 1(3), 31-36.
9. Boerema, G. H. (1993). Contributions towards a monograph of *Phoma* (Coelomycetes)—II. Section *peyronellaea*. *Persoonia-Molecular Phylogeny and Evolution of Fungi*, 15(2), 197-221.
10. Boerema, G. H. (Ed.). (2004). *Phoma identification manual: differentiation of specific and infra-specific taxa in culture*. CABI.
11. Botella, L., & Diez, J. J. (2011). Phylogenetic diversity of fungal endophytes in Spanish stands of *Pinus halepensis*. *Fungal Diversity*, 47, 9-18.
12. Brown, E. F., Bildsten, L., & Rutledge, R. E. (1998). Crustal heating and quiescent emission from transiently accreting neutron stars. *The Astrophysical Journal*, 504(2), L95.

13. Bukharina, I. L., Islamova, N. A., & Lebedeva, M. A. (2018). Species of fungi in the root system of woody plants in urban plantations. *KnE Life Sciences*, 49-55.
14. Chauhan, N. M., Gutama, A. D., & Aysa, A. (2019). Endophytic fungal diversity isolated from different agro-ecosystem of Enset (*Ensete ventericosum*) in Gedeo zone, SNNPRS, Ethiopia. *BMC microbiology*, 19, 1-10.
15. Chen, Z. M., Chen, H. P., Li, Y., Feng, T., & Liu, J. K. (2015). Cytochalasins from cultures of endophytic fungus *Phoma multirostrata* EA-12. *The Journal of Antibiotics*, 68(1), 23-26.
16. Chen, Y., Yang, W., Zou, G., Chen, S., Pang, J., & She, Z. (2019). Bioactive polyketides from the mangrove endophytic fungi *Phoma* sp. SYSU-SK-7. *Fitoterapia*, 139, 104369.
17. Chen, H. M., Wu, H. X., He, X. Y., Zhang, H. H., Miao, F., & Liang, Z. S. (2020). Promoting tanshinone synthesis of *Salvia miltiorrhiza* root by a seed endophytic fungus, *Phoma herbarum* D603. *Zhongguo Zhong yao za zhi= Zhongguo Zhongyao Zazhi= China Journal of Chinese Materia Medica*, 45(1), 65-71.
18. Cosoveanu, A., Gimenez-Mariño, C., Cabrera, Y., Hernandez, G., & Cabrera, R. (2014). Endophytic fungi from grapevine cultivars in Canary Islands and their activity against phytopatogenic fungi. *International Journal of Agriculture and Crop Sciences*, 7(15), 1497.
19. Cosoveanu, A., Rodriguez Sabina, S., & Cabrera, R. (2018). Fungi as endophytes in *Artemisia thuscula*: Juxtaposed elements of diversity and phylogeny. *Journal of Fungi*, 4(1), 17.
20. Costa, I. P., Maia, L. C., & Cavalcanti, M. A. (2012). Diversity of leaf endophytic fungi in mangrove plants of northeast Brazil. *Brazilian Journal of Microbiology*, 43, 1165-1173.
21. Crous, P. W., Petrini, O., Marais, G. F., Pretorius, Z. A., & Rehder, F. (1995). Occurrence of fungal endophytes in cultivars of *Triticum aestivum* in South Africa. *Mycoscience*, 36(1), 105-111.
22. Cui, J. L., Guo, S. X., & Xiao, P. G. (2011). Antitumor and antimicrobial activities of endophytic fungi from medicinal parts of *Aquilaria sinensis*. *Journal of Zhejiang University. Science. B*, 12(5), 385.
23. Das, N., & Chatterjee, T. (2023). *Isolation and Identification of Endophytic Fungi of Rauwolfia Serpentina (L.) Benth. Ex Kurz by DNA Barcoding* (No. 9900). EasyChair.
24. Deshmukh, S. K., Mishra, P. D., Kulkarni-Almeida, A., Verekar, S., Sahoo, M. R., Periyasamy, G., ... & Vishwakarma, R. (2009). Anti-inflammatory and anticancer activity of ergoflavin isolated from an endophytic fungus. *Chemistry & biodiversity*, 6(5), 784-789.

25. De Abreu, L. M., Almeida, A. R., Salgado, M., & Pfenning, L. H. (2010). Fungal endophytes associated with the mistletoe *Phoradendron perrottetii* and its host tree *Tapirira guianensis*. *Mycological Progress*, 9, 559-566.
26. De Oliveira Chagas, M. B., Dos Santos, I. P., da Silva, L. C. N., dos Santos Correia, M. T., de Araújo, J. M., da Silva Cavalcanti, M., & de Menezes Lima, V. L. (2017). Antimicrobial activity of cultivable endophytic fungi associated with *Hancornia speciosa* gomes bark. *The Open Microbiology Journal*, 11, 179.
27. De Vries, S., von Dahlen, J. K., Schnake, A., Ginschel, S., Schulz, B., & Rose, L. E. (2018). Broad-spectrum inhibition of *Phytophthora infestans* by fungal endophytes. *FEMS Microbiology Ecology*, 94(4), fiy037.
28. Douanla-Meli, C., Langer, E., & Mouafo, F. T. (2013). Fungal endophyte diversity and community patterns in healthy and yellowing leaves of *Citrus limon*. *Fungal Ecology*, 6(3), 212-222.
29. Du, W., Yao, Z., Li, J., Sun, C., Xia, J., Wang, B., ... & Ren, L. (2020). Diversity and antimicrobial activity of endophytic fungi isolated from *Securinega suffruticosa* in the Yellow River Delta. *PloS one*, 15(3), e0229589.
30. Ek-Ramos, M. J., Zhou, W., Valencia, C. U., Antwi, J. B., Kalns, L. L., Morgan, G. D., ... & Sword, G. A. (2013). Spatial and temporal variation in fungal endophyte communities isolated from cultivated cotton (*Gossypium hirsutum*). *PLoS One*, 8(6), e66049.
31. El-Nagerabi, S. A., Elshafie, A. E., & Alkhanjari, S. S. (2014). Endophytic fungi associated with endogenous *Boswellia sacra*. *Biodiversitas Journal of Biological Diversity*, 15(1).
32. EL-NAGERABI, S. A., Elshafie, A. E., & Alkhanjari, S. S. (2013). Endophytic fungi associated with *Ziziphus* species and new records from mountainous area of Oman. *Biodiversitas Journal of Biological Diversity*, 14(1).
33. Fernandes, M. D. R. V., Silva, T. A. C., Pfenning, L. H., Costa-Neto, C. M. D., Heinrich, T. A., Alencar, S. M. D., ... & Ikegaki, M. (2009). Biological activities of the fermentation extract of the endophytic fungus *Alternaria alternata* isolated from *Coffea arabica* L. *Brazilian journal of pharmaceutical sciences*, 45, 677-685.
34. Fernandes, E. G., Pereira, O. L., da Silva, C. C., Bento, C. B. P., & de Queiroz, M. V. (2015). Diversity of endophytic fungi in *Glycine max*. *Microbiological Research*, 181, 84-92.

35. Fisher, P. J., Petrini, O., Petrini, L. E., & Sutton, B. C. (1994). Fungal endophytes from the leaves and twigs of *Quercus ilex* L. from England, Majorca and Switzerland. *New Phytologist*, 127(1), 133-137.
36. Gama, D. D. S., Santos, I. A. F. M., Abreu, L. M. D., Medeiros, F. H. V. D., Duarte, W. F., & Cardoso, P. G. (2019). Endophytic fungi from *Brachiaria* grasses in Brazil and preliminary screening of *Sclerotinia sclerotiorum* antagonists. *Scientia Agricola*, 77.
37. García, A., Rhoden, S. A., Rubin Filho, C. J., Nakamura, C. V., & Pamphile, J. A. (2012). Diversity of foliar endophytic fungi from the medicinal plant *Sapindus saponaria* L. and their localization by scanning electron microscopy. *Biological Research*, 45(2), 139-148.
38. García, E., Alonso, Á., Platas, G., & Sacristán, S. (2013). The endophytic mycobiota of *Arabidopsis thaliana*. *Fungal Diversity*, 60, 71-89.
39. Gazis, R., & Chaverri, P. (2010). Diversity of fungal endophytes in leaves and stems of wild rubber trees (*Hevea brasiliensis*) in Peru. *fungal ecology*, 3(3), 240-254.
40. Garibaldi, A., Gilardi, G., Bertetti, D., & Gullino, M. L. (2007). First report of leaf spot and root rot caused by *Phoma betae* on *Beta vulgaris* subsp. *vulgaris* (Garden beet Group) in Italy. *Plant Disease*, 91(11), 1515-1515.
41. Ghimire, S. R., Charlton, N. D., Bell, J. D., Krishnamurthy, Y. L., & Craven, K. D. (2011). Biodiversity of fungal endophyte communities inhabiting switchgrass (*Panicum virgatum* L.) growing in the native tallgrass prairie of northern Oklahoma. *Fungal Diversity*, 47, 19-27.
42. González, V., & Tello, M. L. (2011). The endophytic mycota associated with *Vitis vinifera* in central Spain. *Fungal diversity*, 47(1), 29-42.
43. Gupta, S., Kaul, S., Singh, B., Vishwakarma, R. A., & Dhar, M. K. (2016). Production of gentisyl alcohol from *Phoma herbarum* endophytic in *Curcuma longa* L. and its antagonistic activity towards leaf spot pathogen *Colletotrichum gloeosporioides*. *Applied biochemistry and biotechnology*, 180, 1093-1109.
44. Guo, L. D., Huang, G. R., & Wang, Y. (2008). Seasonal and tissue age influences on endophytic fungi of *Pinus tabulaeformis* (Pinaceae) in the Dongling Mountains, Beijing. *Journal of integrative plant biology*, 50(8), 997-1003.
45. Hamzah, T. N. T., Lee, S. Y., Hidayat, A., Terhem, R., Faridah-Hanum, I., & Mohamed, R. (2018). Diversity and characterization of endophytic fungi isolated from the tropical mangrove species,

- Rhizophora mucronata, and identification of potential antagonists against the soil-borne fungus, *Fusarium solani*. *Frontiers in microbiology*, 9, 1707.
46. Hemamalini, V., Kumar, D. M., Rebecca, A. I. N., Srimathi, S., Muthumary, J., & Kalaichelvan, P. (2015). Isolation and characterization of taxol producing endophytic Phoma sp. from Calotropis gigantea and its anti-proliferative studies. *J. Acad. Ind. Res*, 3, 645-649.
47. Hussain, H., Kock, I., Al-Harrasi, A., Al-Rawahi, A., Abbas, G., Green, I. R., ... & Krohn, K. (2014). Antimicrobial chemical constituents from endophytic fungus Phoma sp. *Asian Pacific Journal of Tropical Medicine*, 7(9), 699-702.
48. Iqbal, S., & Chohan, J. S. (1984). Phoma jolyana, a new pathogen on neem (*Azadirachta indica*). *Indian Forester*, 110(10), 1058-1060.
49. Jayatilake, P. L., & Munasinghe, H. (2020). Antimicrobial activity of cultivable endophytic and rhizosphere fungi associated with “mile-a-minute,” Mikania cordata (Asteraceae). *BioMed Research International*, 2020.
50. Jiaojiao, S., Wattanachai, P., & Kasem, S. (2016). Isolation and identification of endophytic fungi from 10 species palm trees. *Journal of Agricultural Technology*, 12(2), 349-363.
51. Junker, C., Draeger, S., & Schulz, B. (2012). A fine line—endophytes or pathogens in *Arabidopsis thaliana*. *Fungal Ecology*, 5(6), 657-662.
52. Juyal, P., Srivastava, V., & Mathur, A. (2017). Antimicrobial activity of Endophytes from aerial and non aerial parts of *Calotropis procera* against Pathogenic microbes. *Int J Scient Res Pub*, 7(7), 590-596.
53. Kandasamy, P., Manogaran, S., Dhakshinamoorthy, M., & Kannan, K. P. (2015). Evaluation of antioxidant and antibacterial activities of endophytic fungi isolated from *Bauhinia racemosa* Lam and *Phyllanthus amarus* Schum and Thonn. *J Chem Pharm Res*, 7(9), 366-379.
54. Kannan, K. P., Abdul Basheed, M. I., Kannadhasan, S., Pondurai, S., & Dhakshinamoorthy, M. (2017). Mycoendophytes isolated from *Mimusops elengi*. L-A first report. *International Biological and Biomedical Journal*, 3(1), 25-29.
55. Khan, R., Shahzad, S., Choudhary, M. I., Khan, S. A., & Ahmad, A. (2007). Biodiversity of the endophytic fungi isolated from *Calotropis procera* (Ait.) R. Br. *Pakistan Journal of Botany*, 39(6), 2233-2239.

56. Khan, A. L., Waqas, M., Hussain, J., Al-Harrasi, A., Al-Rawahi, A., Al-Hosni, K., ... & Lee, I. J. (2014). Endophytes Aspergillus caespitosus LK12 and Phoma sp. LK13 of Moringa peregrina produce gibberellins and improve rice plant growth. *Journal of Plant Interactions*, 9(1), 731-737.
57. Khan, A. L., Al-Harrasi, A., Al-Rawahi, A., Al-Farsi, Z., Al-Mamari, A., Waqas, M., ... & Lee, I. J. (2016). Endophytic fungi from frankincense tree improves host growth and produces extracellular enzymes and indole acetic acid. *PloS one*, 11(6), e0158207.
58. Kedar, A., Rathod, D., Yadav, A., Agarkar, G., & Rai, M. (2014). Endophytic Phoma sp. isolated from medicinal plants promote the growth of Zea mays. *Nusantara Bioscience*, 6(2).
59. Khiralla, A., Mohamed, I. E., Tzanova, T., Schohn, H., Slezack-Deschaumes, S., Hehn, A., ... & Laurain-Mattar, D. (2016). Endophytic fungi associated with Sudanese medicinal plants show cytotoxic and antibiotic potential. *FEMS microbiology letters*, 363(11).
60. Kim, J. W., Ko, W., Kim, E., Kim, G. S., Hwang, G. J., Son, S., ... & Ahn, J. S. (2018). Anti-inflammatory phomalichenones from an endolichenic fungus Phoma sp. *The Journal of Antibiotics*, 71(8), 753-756.
61. Kumar, S., Upadhyay, R., Aharwal, R. P., & Sandhu, S. S. (2016). Antibacterial activity of some isolated endophytic fungi from Menthe viridis. *Int J Appl Biol Pharm Technol*, 7, 239-248.
62. Kumaran, R. S., Choi, Y. K., Lee, S., Jeon, H. J., Jung, H., & Kim, H. J. (2012). Isolation of taxol, an anticancer drug produced by the endophytic fungus, Phoma betae. *African Journal of Biotechnology*, 11(4), 950-960.
63. Krishnamurthy, Y. L., Naik, S. B., & Jayaram, S. (2008). Fungal communities in herbaceous medicinal plants from the Malnad region, Southern India. *Microbes and environments*, 23(1), 24-28.
64. Kwong-Chung, K.J.; Bennett, J.E. (1992). Medical Mycology; Lea & Febiger: Philadelphia, PA, USA, pp. 661–662, ISBN 0812114639
65. Larrán, S., Perello, A., Simón, M. R., & Moreno, V. (2002). Isolation and analysis of endophytic microorganisms in wheat (*Triticum aestivum* L.) leaves. *World Journal of Microbiology and Biotechnology*, 18, 683-686.
66. Li, H. T., Liu, T., Yang, R., Xie, F., Yang, Z., Yang, Y., ... & Ding, Z. T. (2020). Phomretones A–F, C 12 polyketides from the co-cultivation of Phoma sp. YUD17001 and Armillaria sp. *RSC advances*, 10(31), 18384-18389.

67. Li, X., Zhai, X., Shu, Z., Dong, R., Ming, Q., Qin, L., & Zheng, C. (2016). *Phoma glomerata* D14: An endophytic fungus from *Salvia miltiorrhiza* that produces salvianolic acid C. *Current Microbiology*, 73, 31-37.
68. Liu, S. S., Jiang, J. X., Huang, R., Wang, Y. T., Jiang, B. G., Zheng, K. X., & Wu, S. H. (2019). A new antiviral 14-nordrimane sesquiterpenoid from an endophytic fungus *Phoma* sp. *Phytochemistry Letters*, 29, 75-78.
69. Luiz H. R., Nurhayat, T., Zhiqiang, P., Ulrich R. B., James J. B., Natasha M. A., ... & Rita M. M. (2012). Diversity and Biological Activities of Endophytic Fungi Associated with Micropropagated Medicinal Plant *Echinacea purpurea* (L.) Moench. *American Journal of Plant Sciences*, 2012.
70. Loro, M., Valero-Jiménez, C. A., Nozawa, S., & Márquez, L. M. (2012). Diversity and composition of fungal endophytes in semiarid Northwest Venezuela. *Journal of Arid Environments*, 85, 46-55.
71. Lv, R., Zheng, L., Zhu, Z., Pan, L., Huang, J., & Hsiang, T. (2011). First report of stem blight of *Eleocharis dulcis* caused by *Phoma bellidis* in China. *Plant Disease*, 95(9), 1190-1190.
72. Macia-Vicente, J. G., Jansson, H. B., Mendgen, K., & Lopez-Llorca, L. V. (2008). Colonization of barley roots by endophytic fungi and their reduction of take-all caused by *Gaeumannomyces graminis* var. *tritici*. *Canadian Journal of Microbiology*, 54(8), 600-609.
73. Maheswari, S., & Rajagopal, K. (2013). Biodiversity of endophytic fungi in *Kigelia pinnata* during two different seasons. *Current Science*, 515-518.
74. Masumi, S., Mirzaei, S., Kalvandi, R., & Zafari, D. (2014). Asparaginase and amylase activity of thyme endophytic fungi. *Journal of Crop Protection*, 3(5), 655-662.
75. Materatski, P., Varanda, C., Carvalho, T., Dias, A. B., Campos, M. D., Rei, F., & do Rosário Félix, M. (2019). Spatial and temporal variation of fungal endophytic richness and diversity associated to the phyllosphere of olive cultivars. *Fungal biology*, 123(1), 66-76.
76. Moricca, S., Ginetti, B., & Ragazzi, A. (2012). Species-and organ-specificity in endophytes colonizing healthy and declining Mediterranean oaks. *Phytopathologia Mediterranea*, 587-598.
77. Myrchiang, P., Dkhar, M. S., & Devi, H. R. (2014). Studies on endophytic fungi associated with medicinally important aromatic plant *Artemisia nilagirica* (CB Clarke) Pamp. and their antagonistic activity against *Phytophthora infestans*. *J. Adv. Lab. Res. Biol.*, 5, 112-119.
78. Nalli, Y., Arora, P., Khan, S., Malik, F., Riyaz-Ul-Hassan, S., Gupta, V., & Ali, A. (2019). Isolation, structural modification of macrophin from endophytic fungus *Phoma macrostoma* and their cytotoxic potential. *Medicinal Chemistry Research*, 28, 260-266.

79. Newsham, K. K. (1994). First record of intracellular sporulation by a coelomycete fungus. *Mycological Research*, 98(12), 1390-1392.
80. Novak Babič, M., Gunde-Cimerman, N., Vargha, M., Tischner, Z., Magyar, D., Veríssimo, C., ... & Brandão, J. (2017). Fungal contaminants in drinking water regulation? A tale of ecology, exposure, purification and clinical relevance. *International journal of environmental research and public health*, 14(6), 636.
81. Novas, M. V., & Carmarán, C. C. (2008). Studies on diversity of foliar fungal endophytes of naturalised trees from Argentina, with a description of *Haplotrichum minutissimum* sp. nov. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 203(7), 610-616.
82. Oliveira, R. J. V., Souza, R. G., Lima, T. E. F., & Cavalcanti, M. A. Q. (2014). Endophytic fungal diversity in coffee leaves (*Coffea arabica*) cultivated using organic and conventional crop management systems. *Mycosphere*, 5(4), 523-530.
83. Orlandelli, R. C., Alberto, R. N., Rubin Filho, C. J., & Pamphile, J. A. (2012). Diversity of endophytic fungal community associated with *Piper hispidum* (Piperaceae) leaves. *Genet Mol Res*, 11(2), 1575-1585.
84. Osterhage, C., König, G. M., Jones, P. G., & Wright, A. D. (2002). 5-Hydroxyramulosin, a new natural product produced by *Phoma tropica*, a marine-derived fungus isolated from the alga *Fucus spiralis*. *Planta medica*, 68(11), 1052-1054.
85. Parismita, S., Manoj, K., Highland, K., Ruth, L., & Richa, R. (2016). Endophytic fungi associated with the medicinally important aromatic plant *Artemisia nilagirica* (Clarke) Pamp. and antimicrobial activity of selected endophytic fungi against *Rhizoctonia solani*. *Asian Journal of Biological and Life Science*, 5(2).
86. Park, S. U., Lim, H. S., Park, K. C., Park, Y. H., & Bae, H. (2012). Fungal endophytes from three cultivars of *Panax ginseng* Meyer cultivated in Korea. *Journal of ginseng research*, 36(1), 107.
87. Park, Y. H., Chung, J. Y., Ahn, D. J., Kwon, T. R., Lee, S. K., Bae, I., ... & Bae, H. (2015). Screening and characterization of endophytic fungi of *Panax ginseng* Meyer for biocontrol activity against ginseng pathogens. *Biological Control*, 91, 71-81.
88. Paul, N. C., Kim, W. K., Woo, S. K., Park, M. S., & Yu, S. H. (2006). Diversity of endophytic fungi associated with *Taraxacum coreanum* and their antifungal activity. *Mycobiology*, 34(4), 185-190.

89. Pharamat, T., Palaga, T., Piapukiew, J., Whalley, A. J., & Sihanonth, P. (2013). Antimicrobial and anticancer activities of endophytic fungi from *Mitragyna javanica* Koord and Val. *Afr J Microbiol Res*, 7(49), 5565-5572.
90. Poveda, J., Zabalgogeazcoa, I., Soengas, P., Rodríguez, V. M., Cartea, M. E., Abilleira, R., & Velasco, P. (2020). Brassica oleracea var. acephala (kale) improvement by biological activity of root endophytic fungi. *Scientific Reports*, 10(1), 20224.
91. Purmale, L., Apine, I., Nikolajewa, V., Grantina, L., Verkley, G., & Tomsone, S. (2012). Endophytic fungi in evergreen rhododendron cultivated in vitro and in vivo. *Environmental and Experimental Biology*, 10, 1-7.
92. RADIASTUTI, N., SUSILOWATI, D. N., & BAHALWAN, H. A. (2019). Phylogenetic study of endophytic fungi associated with *Centella asiatica* from Bengkulu and Malaysian accessions based on the ITS rDNA sequence. *Biodiversitas Journal of Biological Diversity*, 20(5).
93. Ragazzi, A., Moricca, S., Capretti, P., & Dellavalle, I. (1999). Endophytic presence of *Discula quercina* on declining *Quercus cerris*. *Journal of Phytopathology*, 147(7-8), 437-440.
94. Rai, M. K., Tiwari, V. V., Irinyi, L., & Kövics, G. J. (2014). Advances in taxonomy of genus *Phoma*: polyphyletic nature and role of phenotypic traits and molecular systematics. *Indian journal of microbiology*, 54, 123-128.
95. Rajak, R. C., & Rai, M. K. (1983). Effect of different factors on the morphology and cultural characters of 18-species and 5-varieties of *Phoma*. I. Effect of different media. *Bibliotheca Mycologica*, 91, 301-317.
96. Rajagopal, K., Mahendran, T. S., & Selvakumar, S. (2008). Preliminary studies on the diversity of the fungal endophytes in few *Ficus* Species. *Indian Journal of Applied Microbiology*, 8(1), 11-14.
97. Rivera-Orduña, F. N., Suarez-Sánchez, R. A., Flores-Bustamante, Z. R., Gracida-Rodriguez, J. N., & Flores-Cotera, L. B. (2011). Diversity of endophytic fungi of *Taxus globosa* (Mexican yew). *Fungal Diversity*, 47, 65-74.
98. Sánchez Márquez, S., Bills, G. F., Domínguez Acuña, L., & Zabalgogeazcoa, I. (2010). Endophytic mycobiota of leaves and roots of the grass *Holcus lanatus*. *Fungal Diversity*, 41, 115-123.
99. Sandhu, S. S., Kumar, S., & Aharwal, R. P. (2014). Isolation and identification of endophytic fungi from *Ricinus communis* Linn. and their antibacterial activity. *Int. J. Res. Pharm. Chem*, 4(3), 611-618.

100. Sánchez Márquez, S., Bills, G. F., Domínguez Acuña, L., & Zabalgogeazcoa, I. (2010). Endophytic mycobiota of leaves and roots of the grass *Holcus lanatus*. *Fungal Diversity*, 41, 115-123.
101. SCHULZ, B., GUSKE, S., & DAMMANN, U. (1998). Endophyte-host interactions. II. Defining symbiosis of the endophyte-host interaction. *Symbiosis*.
102. Selim, K. A., Elkhateeb, W. A., Tawila, A. M., El-Beih, A. A., Abdel-Rahman, T. M., El-Diwany, A. I., & Ahmed, E. F. (2018). Antiviral and antioxidant potential of fungal endophytes of Egyptian medicinal plants. *Fermentation*, 4(3), 49.
103. Selvanathan, S., Indrakumar, I., & Johnpaul, M. (2011). Biodiversity of the endophytic fungi isolated from *Calotropis gigantea* (L.) R. Br. *Recent Research in Science and Technology*, 3(4).
104. Sim, J. H., Khoo, C. H., Lee, L. H., & Cheah, Y. K. (2010). Molecular diversity of fungal endophytes isolated from *Garcinia mangostana* and *Garcinia parvifolia*. *J Microbiol Biotechnol*, 20(4), 651-658.
105. Singh, G., Katoch, A., Razak, M., Kitchlu, S., Goswami, A., & Katoch, M. (2017). Bioactive and biocontrol potential of endophytic fungi associated with *Brugmansia aurea* Lagerh. *FEMS Microbiology Letters*, 364(21).
106. Sinneto, S., Alonso, R., Tiscornia, S., & Bettucci, I. (2005). Fungal community of *Eucalyptus globulus* and *Eucalyptus maidenii* stems in Uruguay.
107. Smith, H., Wingfield, M. J., & Petrini, O. (1996). Botryosphaeria dothidea endophytic in *Eucalyptus grandis* and *Eucalyptus nitens* in South Africa. *Forest ecology and management*, 89(1-3), 189-195.
108. Song, J., Wattanachai, P., & Kasem, S. (2015). Biological activity of endophytic fungi associated with palm trees. *Journal of Agricultural Technology*, 11(2), 567-579.
109. Srivastava, A. P. A. R. N. A., & Anandrao, R. K. (2015). Antimicrobial potential of fungal endophytes isolated from leaves of *Prosopis juliflora* (SW.) DC. an important weed. *International Journal of Pharmacy and Pharmaceutical Sciences*, 7(12), 128-136.
110. Strobel, G., Singh, S. K., Riyaz-Ul-Hassan, S., Mitchell, A. M., Geary, B., & Sears, J. (2011). An endophytic/pathogenic *Phoma* sp. from creosote bush producing biologically active volatile compounds having fuel potential. *FEMS microbiology letters*, 320(2), 87-94.

111. Sun, J., Guo, L., Zang, W., Ping, W., & Chi, D. (2008). Diversity and ecological distribution of endophytic fungi associated with medicinal plants. *Science in China Series C: Life Sciences*, 51(8), 751-759.
112. Sun, X., Guo, L. D., & Hyde, K. D. (2011). Community composition of endophytic fungi in *Acer truncatum* and their role in decomposition. *Fungal diversity*, 47, 85-95.
113. Sun, Y., Wang, Q., Lu, X., Okane, I., & Kakishima, M. (2012). Endophytic fungal community in stems and leaves of plants from desert areas in China. *Mycological Progress*, 11, 781-790.
114. Suryan, L., Aruna, A., & Namasivayam, S. (2016). Screening of antibacterial activity of endophytic fungi *Phoma europhyrena* against human pathogenic bacteria. *Research Journal of Pharmacy and Technology*, 9(4), 437-439.
115. Szűcs, Z., Plaszkó, T., Cziáky, Z., Kiss-Szikszai, A., Emri, T., Bertóti, R., ... & Gonda, S. (2018). Endophytic fungi from the roots of horseradish (*Armoracia rusticana*) and their interactions with the defensive metabolites of the glucosinolate-myrosinase-isothiocyanate system. *BMC plant biology*, 18(1), 1-15.
116. Tan, X. M., Li, L. Y., Sun, L. Y., Sun, B. D., Niu, S. B., Wang, M. H., ... & Ding, G. (2018)a. Spiciferone analogs from an endophytic fungus *Phoma betae* collected from desert plants in West China. *The Journal of Antibiotics*, 71(6), 613-617.
117. Tan, X. M., Zhou, Y. Q., Zhou, X. L., Xia, X. H., Wei, Y., He, L. L., ... & Yu, L. Y. (2018)b. Diversity and bioactive potential of culturable fungal endophytes of *Dysosma versipellis*; a rare medicinal plant endemic to China. *Scientific Reports*, 8(1), 5929.
118. Taylor, J. E., Hyde, K. D., & Jones, E. B. G. (1999). Endophytic fungi associated with the temperate palm, *Trachycarpus fortunei*, within and outside its natural geographic range. *The New Phytologist*, 142(2), 335-346.
119. Teimoori-Boghsani, Y., Ganjeali, A., Cernava, T., Müller, H., Asili, J., & Berg, G. (2020). Endophytic fungi of native *Salvia abrotanoides* plants reveal high taxonomic diversity and unique profiles of secondary metabolites. *Frontiers in microbiology*, 10, 3013.
120. Toghueo, R. M. K., Zabalgogeazcoa, I., de Aldana, B. V., & Boyom, F. F. (2017). Enzymatic activity of endophytic fungi from the medicinal plants *Terminalia catappa*, *Terminalia mantaly* and *Cananga odorata*. *South African Journal of Botany*, 109, 146-153.

121. Vieira, P. D. D. S., Motta, C. M. D. S., Lima, D., Torres, J. B., Quecine, M. C., Azevedo, J. L., & Oliveira, N. T. D. (2011). Endophytic fungi associated with transgenic and non-transgenic cotton. *Mycology*, 2(2), 91-97.
122. Vieira, M. L., Hughes, A. F., Gil, V. B., Vaz, A. B., Alves, T. M., Zani, C. L., ... & Rosa, L. H. (2012). Diversity and antimicrobial activities of the fungal endophyte community associated with the traditional Brazilian medicinal plant Solanum cernuum Vell.(Solanaceae). *Canadian journal of microbiology*, 58(1), 54-66.
123. Vujanovic, V., & Brisson, J. (2002). A comparative study of endophytic mycobiota in leaves of Acer saccharum in eastern North America. *Mycological Progress*, 1(2), 147-154.
124. Wang, L. W., Xu, B. G., Wang, J. Y., Su, Z. Z., Lin, F. C., Zhang, C. L., & Kubicek, C. P. (2012). Bioactive metabolites from Phoma species, an endophytic fungus from the Chinese medicinal plant Arisaema erubescens. *Applied microbiology and biotechnology*, 93, 1231-1239.
125. Wang, W. X., Zheng, M. J., Li, J., Feng, T., Li, Z. H., Huang, R., ... & Liu, J. K. (2019). Cytotoxic polyketides from endophytic fungus Phoma bellidis harbored in Tricyrtis maculata. *Phytochemistry Letters*, 29, 41-46.
126. Wati, M. S., & Hadiwiyono, A. Y. (2019). Antagonism of Endophytic Fungi Isolates Artemisia Annua Towards Rhizoctonia Solani, Causal Agent Of Rice Sheath Blight. *International Journal of Innovations in Engineering and Technology*, 14(1), 75-79.
127. Waqas, M., Khan, A. L., Kamran, M., Hamayun, M., Kang, S. M., Kim, Y. H., & Lee, I. J. (2012). Endophytic fungi produce gibberellins and indoleacetic acid and promotes host-plant growth during stress. *Molecules*, 17(9), 10754-10773.
128. Weber, R. W., Stenger, E., Meffert, A., & Matthias, H. A. H. N. (2004). Brefeldin A production by Phoma medicaginis in dead pre-colonized plant tissue: a strategy for habitat conquest?. *Mycological research*, 108(6), 662-671.
129. Wijeratne, E. K., He, H., Franzblau, S. G., Hoffman, A. M., & Gunatilaka, A. L. (2013). Phomapyrrolidones A–C, antitubercular alkaloids from the endophytic fungus Phoma sp. NRRL 46751. *Journal of Natural Products*, 76(10), 1860-1865.
130. Xiong, Z. Q., Yang, Y. Y., Zhao, N., & Wang, Y. (2013). Diversity of endophytic fungi and screening of fungal paclitaxel producer from Anglojap yew, Taxus x media. *BMC microbiology*, 13(1), 1-10.

131. Yang, X., Strobel, G., Stierle, A., Hess, W. M., Lee, J., & Clardy, J. (1994). A fungal endophyte-tree relationship: *Phoma* sp. in *Taxus wallachiana*. *Plant Science*, 102(1), 1-9.
132. Zaferanloo, B., Pepper, S. A., Coulthard, S. A., Redfern, C. P., & Palombo, E. A. (2018). Metabolites of endophytic fungi from Australian native plants as potential anticancer agents. *FEMS microbiology letters*, 365(9), fny078.
133. Zaiyou, J., Li, M., & Xiqiao, H. (2017). An endophytic fungus efficiently producing paclitaxel isolated from *Taxus wallichiana* var. *mairei*. *Medicine*, 96(27).
134. Zakaria, L., & Aziz, W. N. W. (2018). Molecular identification of endophytic fungi from banana leaves (*Musa* spp.). *Tropical life sciences research*, 29(2), 201.
135. Zheng, Y. K., Miao, C. P., Chen, H. H., Huang, F. F., Xia, Y. M., Chen, Y. W., & Zhao, L. X. (2017). Endophytic fungi harbored in *Panax notoginseng*: diversity and potential as biological control agents against host plant pathogens of root-rot disease. *Journal of Ginseng Research*, 41(3), 353-360.